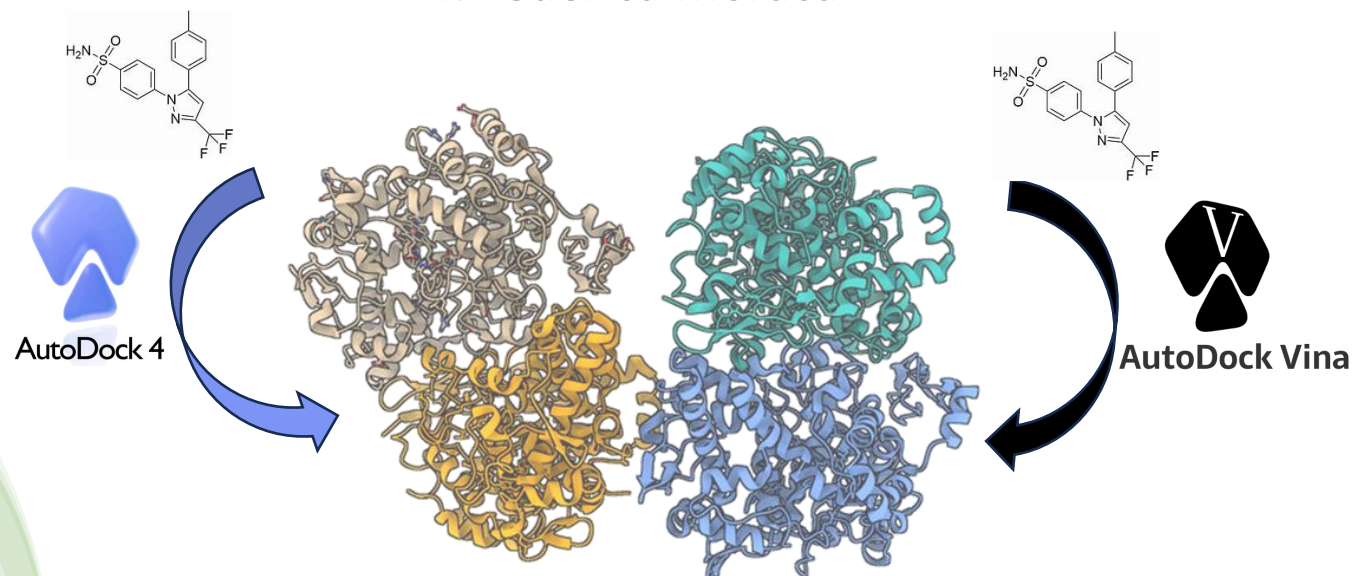


7th Advanced in silico Drug Design workshop/challenge 2024

Molecular Docking Tutorial

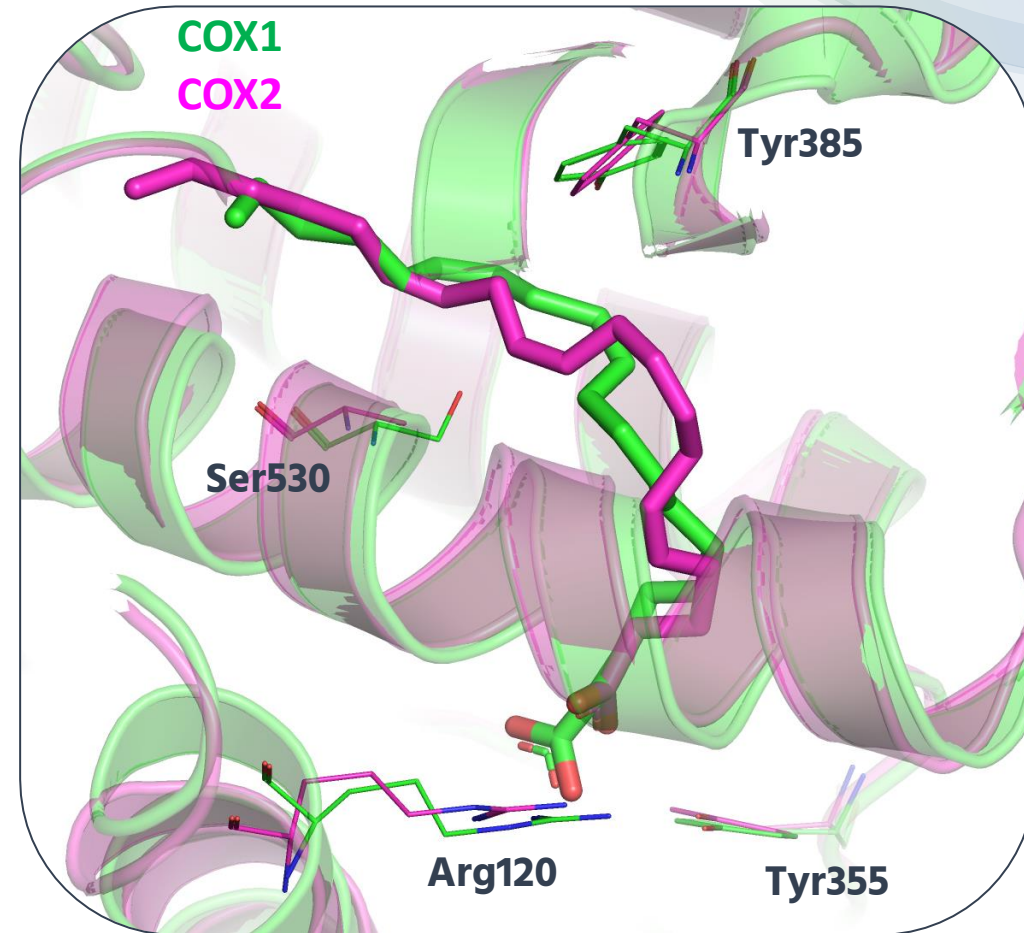
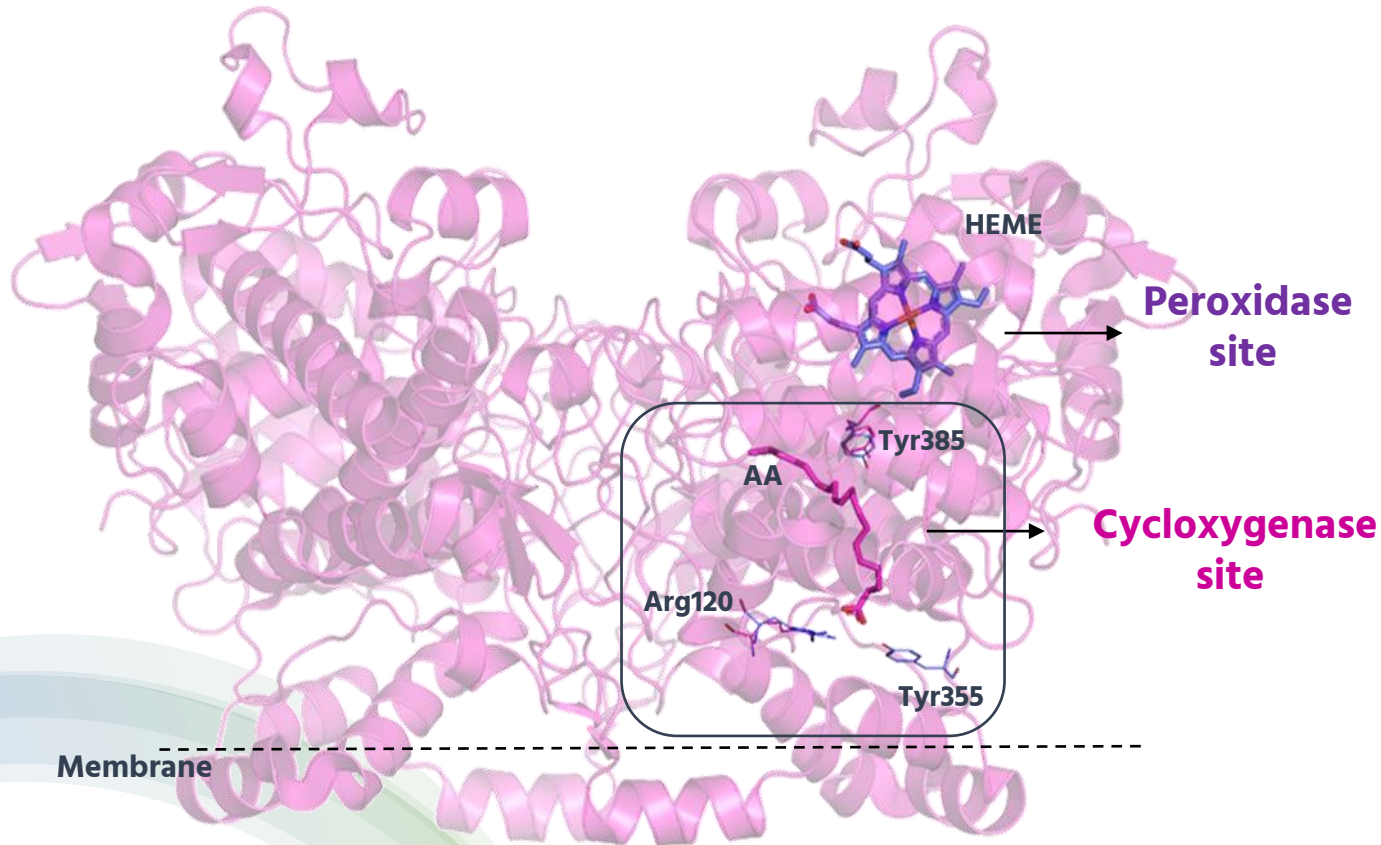
Dr. Federica Moraca



UP Olomouc 29.01. -02.02.2024



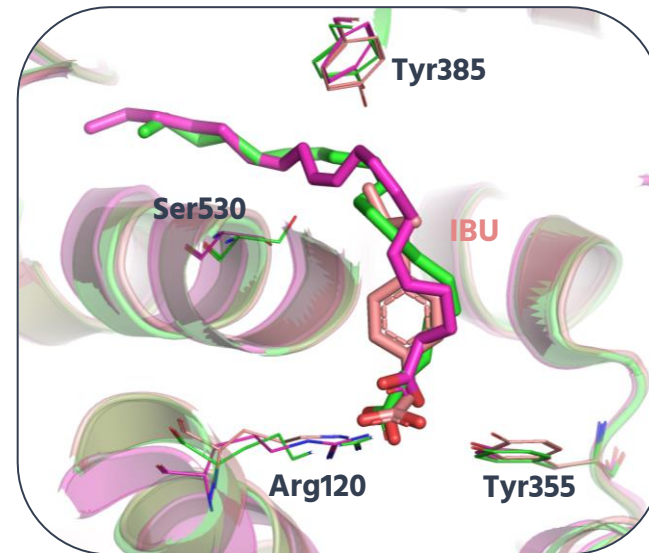
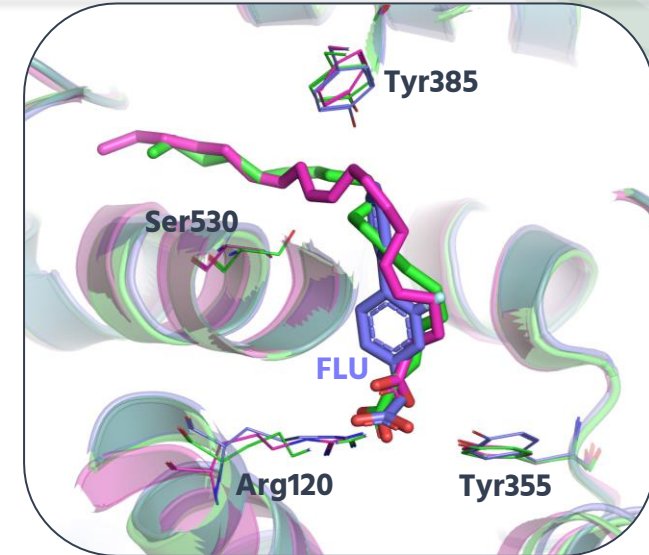
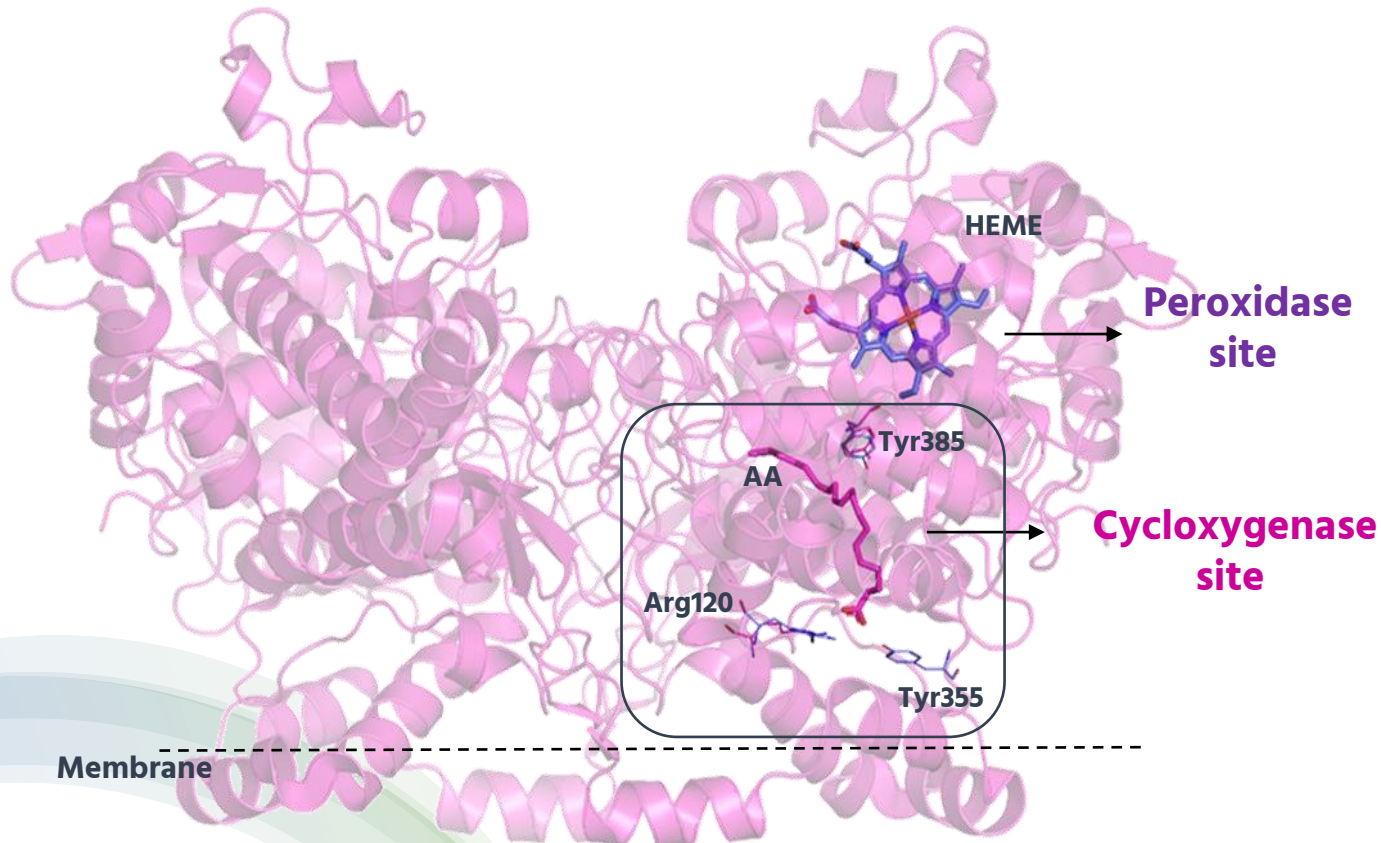
COXs Mechanism of Action



The X-ray structure of COXs complexed with Arachidonic Acid (AA) confirms a L-shaped binding conformation, with the carboxylate moiety of AA binding to **Arg120** and **Tyr355**, while the omega-end positioned in a region termed the top channel in close contact with **Ser530** and **Tyr385**



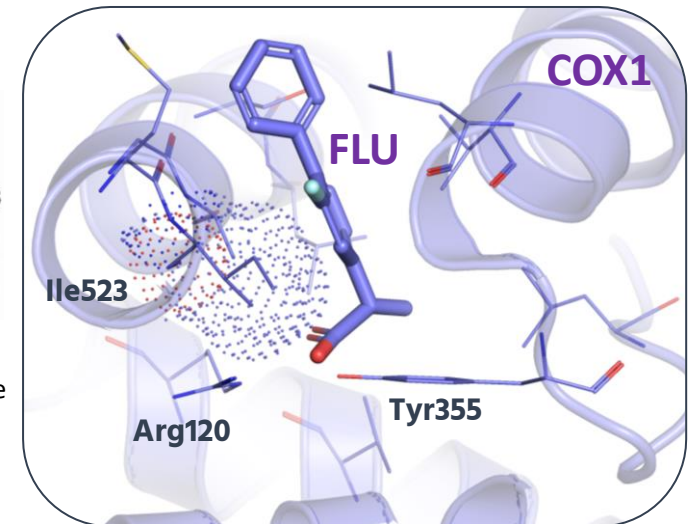
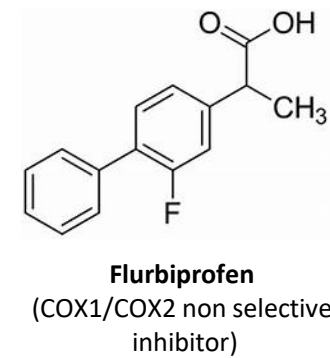
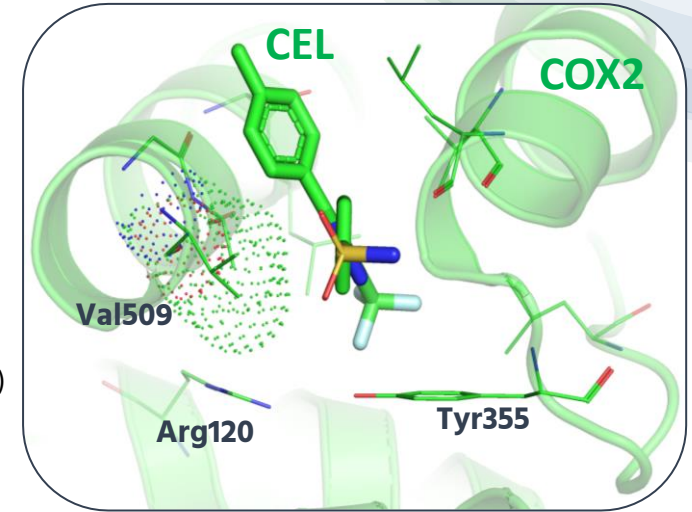
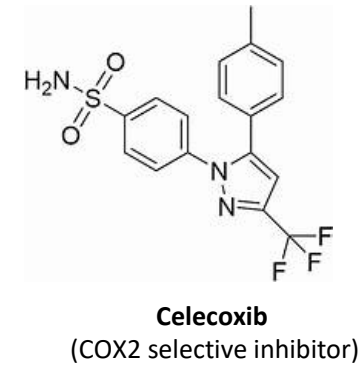
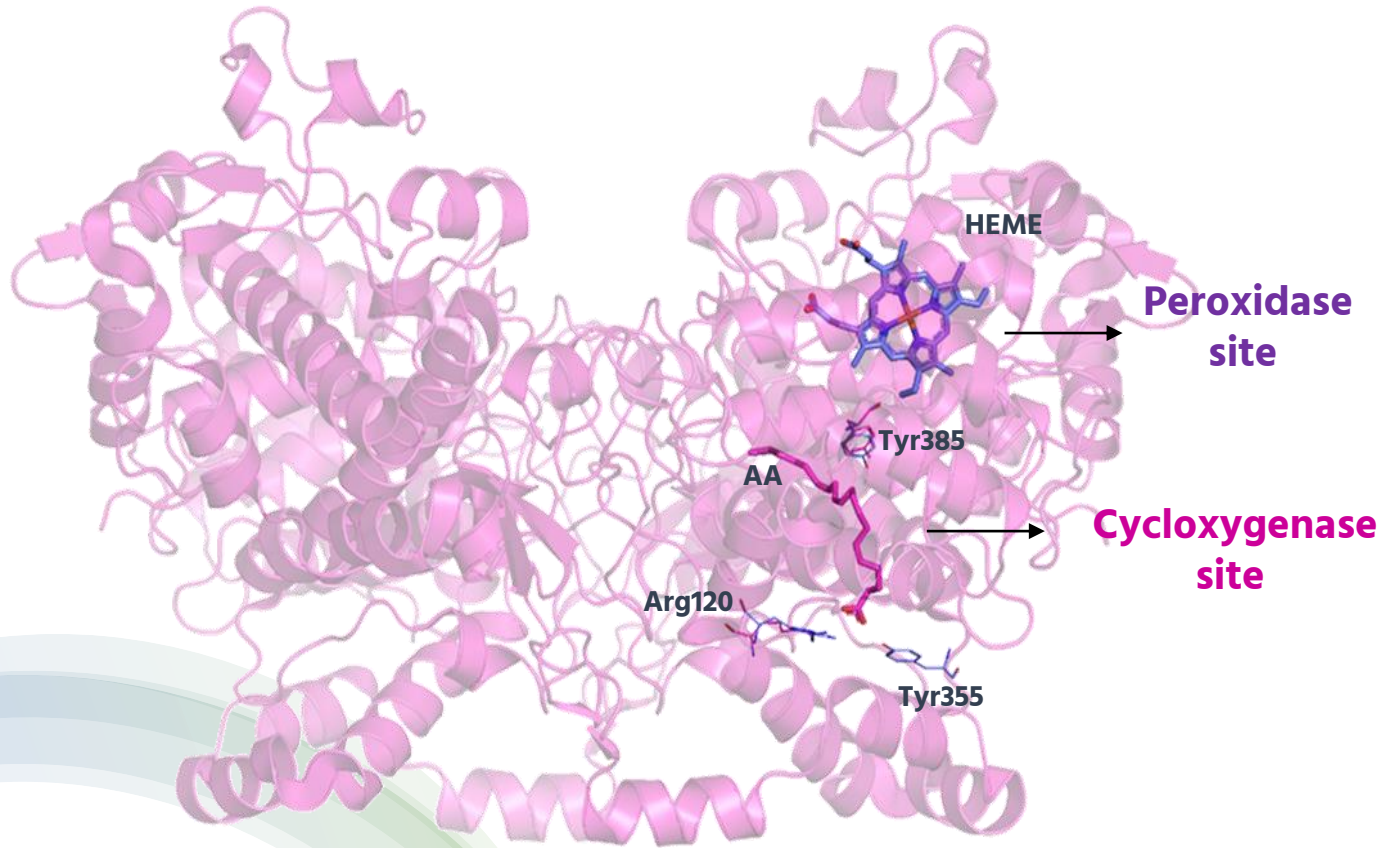
Mechanism of Action of NSAIDs against COXs



Reversible competitive inhibitors (Ibuprofen and Flurbiprofen) act by interfering with hydrophilic interactions (hydrogen-bonds or salt-bridge) with **Arg120** and **Tyr355** at the entrance of the cyclooxygenase channel.



Structural differences between COX1 and COX2



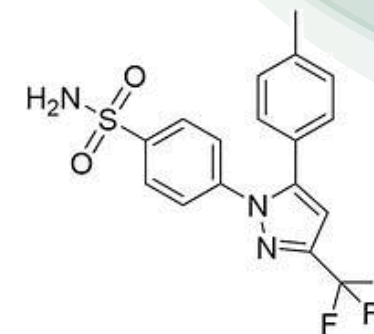


Molecular Docking Tutorial



TASK:

Perform Molecular Docking calculations of Celecoxib against both the COX1 and COX2 isoforms, in order to understand the molecular basis of its COX2 selectivity



Celecoxib
(COX2 selective inhibitor)

- 1. MGLTools (GUI of AutoDock Tools)**
- 2. AutoDock4 and AutoDock Vina docking engines**
- 3. PyMOL (visualization results)**



Molecular Docking Tutorial



AutoDock 4



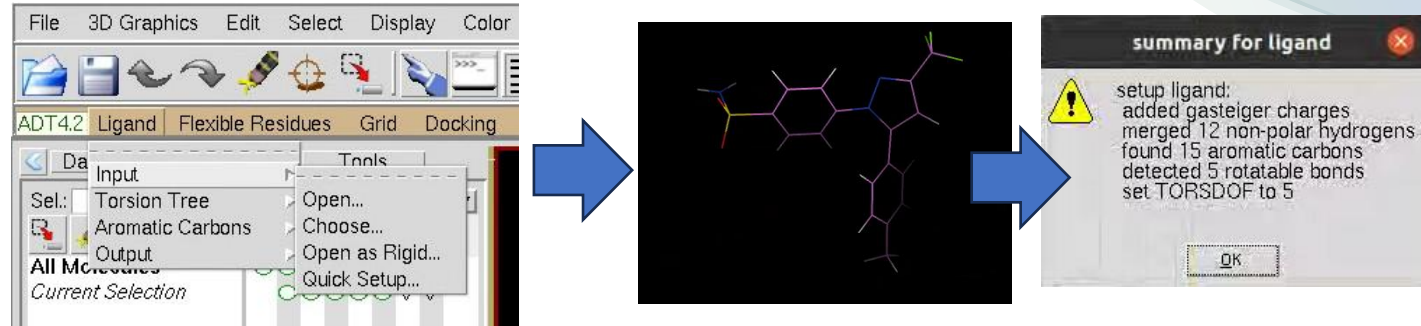
AutoDock 4

Molecular Docking Tutorial

Ligand preparation (GUI of AutoDock Tools)

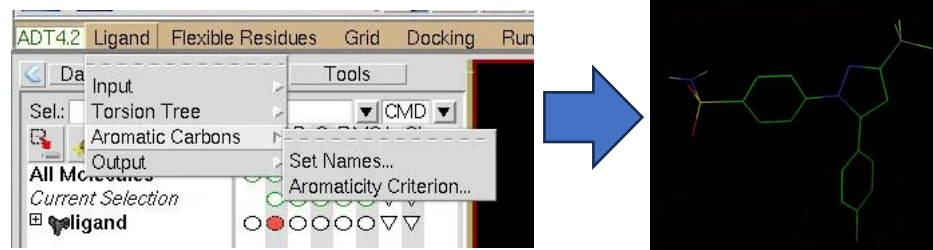
1. Import the Celecoxib (CEL.pdb)

Ligand>Input>Open>*.pdb>CEL.pdb
ligand will be prepared for docking. After clicking the "OK" button, you will see that all the hydrogens atoms are merged to carbon atoms.



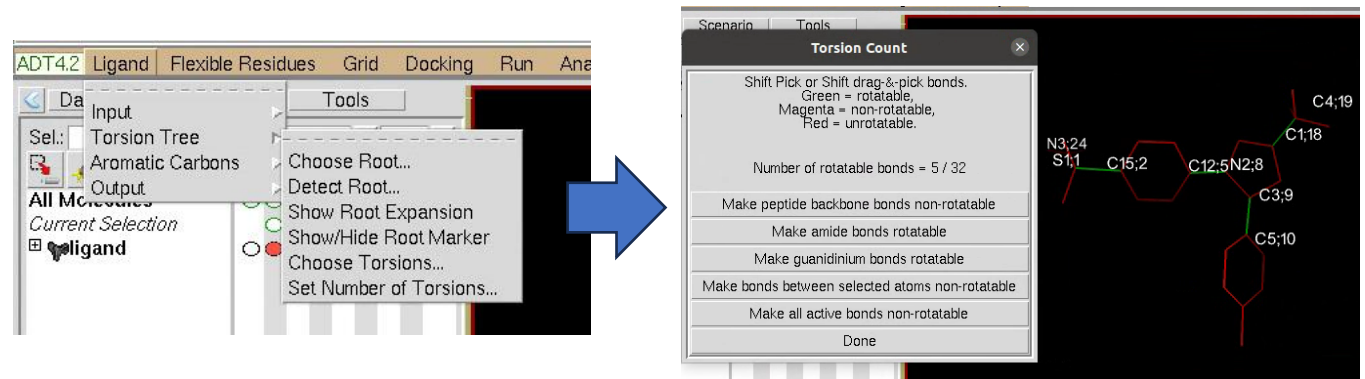
2. Check for aromatic carbons

Ligand>Aromatic carbons>Set Names
Aromatic atoms are shown in green



3. Check rotatable torsions

Ligand>Torsion Tree>Choose Torsion
Rotatable bonds are shown in green, unrotatable in red. Celecoxib has 5 rotatable bonds



4. Save pdbqt file

Ligand>Output>Save as> *.pdbqt>CEL.pdbqt



Molecular Docking Tutorial

Let's have a look on the CEL.pdbqt file

```
REMARK 5 active torsions:
REMARK status: ('A' for Active; 'I' for Inactive)
REMARK 1 A between atoms: S1_1 and C15_2
REMARK 2 A between atoms: S1_1 and N3_24
REMARK 3 A between atoms: C12_5 and N2_8
REMARK 4 A between atoms: C3_9 and C5_10
REMARK 5 A between atoms: C1_18 and C4_19
```

Information about
ligand active (A)
torsions

ROOT					Coor x	Coor y	Coor z	Occ.	B-factor	Charges	Types	
ATOM	1	S1	CEL	A	682	25.931	-21.467	-17.155	1.00	43.78	0.256	S
ATOM	2	O2	CEL	A	682	25.772	-20.039	-17.291	1.00	45.34	-0.201	OA
ATOM	3	O1	CEL	A	682	25.436	-22.106	-15.949	1.00	45.63	-0.201	OA

Atom description

ENDROOT

BRANCH 1 4

Torsion definitions

ATOM	4	C15	CEL	A	682	27.679	-21.706	-17.131	1.00	41.41	0.079	A
ATOM	5	C14	CEL	A	682	28.218	-22.829	-16.556	1.00	39.70	0.027	A
ATOM	6	C13	CEL	A	682	29.584	-22.964	-16.543	1.00	39.68	0.033	A
ATOM	7	C12	CEL	A	682	30.341	-21.967	-17.109	1.00	39.92	0.059	A
ATOM	8	C17	CEL	A	682	29.796	-20.853	-17.697	1.00	40.26	0.033	A
ATOM	9	C16	CEL	A	682	28.434	-20.714	-17.707	1.00	40.98	0.027	A
BRANCH	7	10										
ATOM	10	N2	CEL	A	682	31.724	-22.006	-17.132	1.00	40.07	-0.233	N
ATOM	11	C3	CEL	A	682	32.622	-22.574	-16.260	1.00	40.53	0.071	A
ATOM	12	C2	CEL	A	682	33.843	-22.337	-16.837	1.00	41.52	0.070	A

TORSDOF 5

Number of active torsion

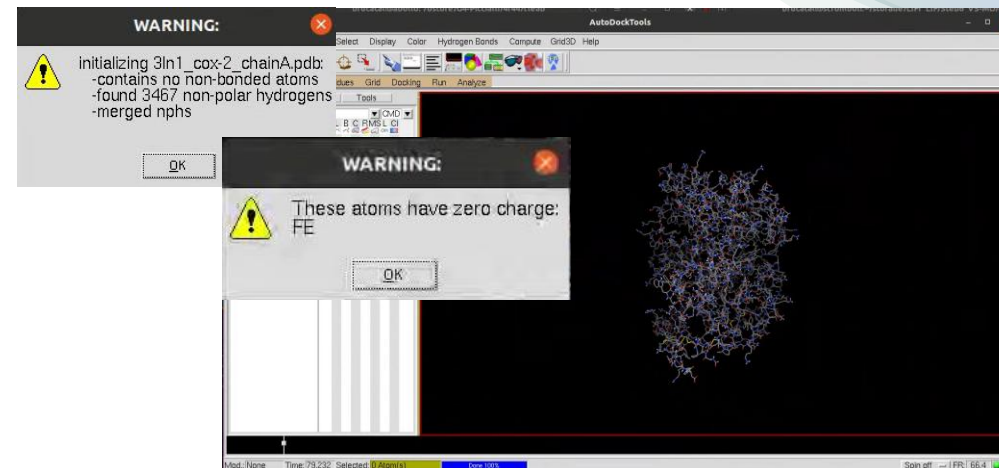
Molecular Docking Tutorial

Receptor preparation (GUI of AutoDock Tools)

1. Select protein (only ChainA)

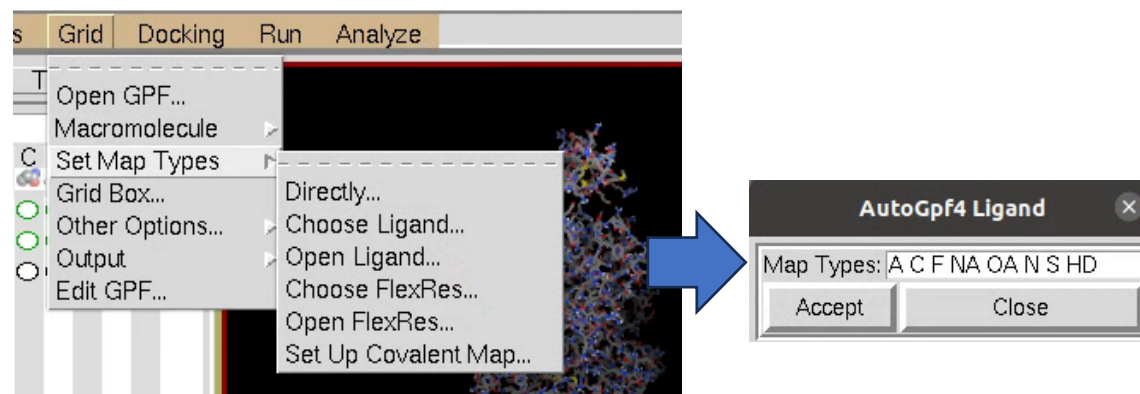
Grid>Macromolecule>Open>cox2.pdb

protein will be prepared for docking (nonpolar hydrogens merged with carbons, charges assigned)



2. Set the Celecoxib Map Types

Grid>Set Map Types>Directly (A C F NA OA N S HD)



COX2 preparation (GUI of AutoDock Tools)

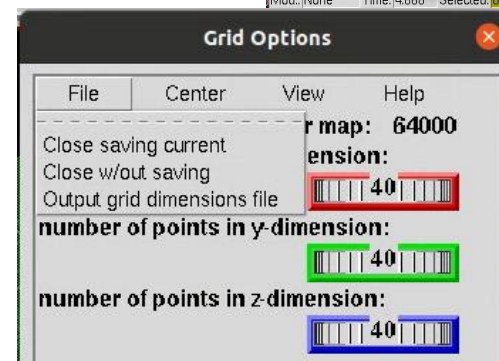
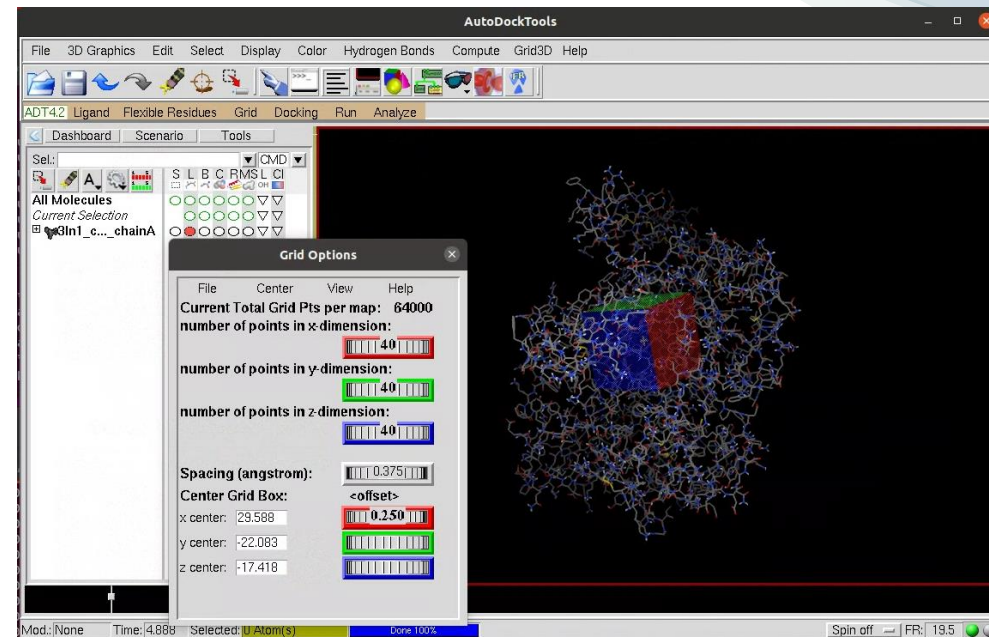
3. Locate the Grid Box on the cyclooxygenase site

Grid>Grid Box

- A cube with a default size of 40x40x40 will appear.
- Adjust the box coordinates so it will cover active site (at the center of CEL), but not much more. For a good centering of the grid box we suggest to manually modify to
x center: 29.588,
y center: -22.083,
z center: -17.418

4. Save the Grid Box Parameter

From the Grid Options window: File> Close saving current




Molecular Docking Tutorial

COX2 preparation (GUI of AutoDock Tools)

5. Manually edit protein PDBQT to charge iron (+2)

```
HETATM 4482 FE      HEM A 500          5.890  24.568  -1.058  1.00 18.80      2.000 Fe
```

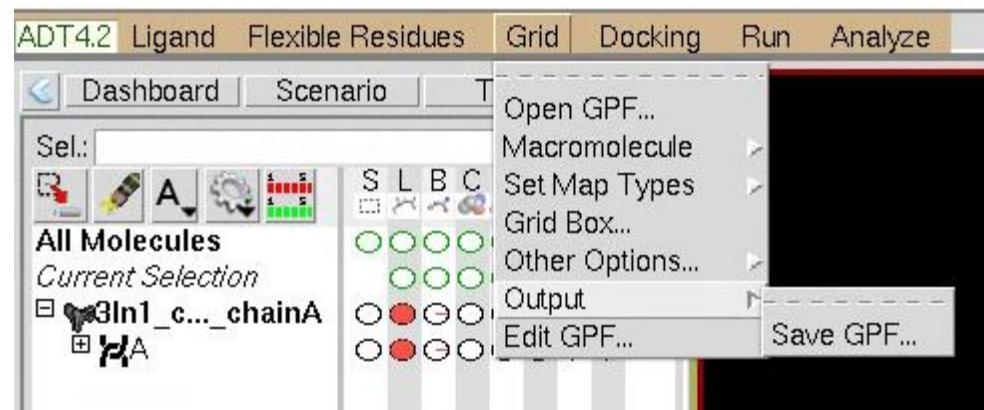


6. Save the Grid Parameter File (GPF)

```
Grid>Output>Save GPF>cox-2.gpf
```

Alternatively you can run the following command

```
prepare_gpf4.py -l CEL.pdbqt -r cox2.pdbqt -o cox-2.gpf
```



7. Run AutoGrid

```
autogrid4 -p cox-2.gpf -l cox2.glg
```

Repeat the same procedure for COX1...

Molecular Docking Tutorial

Preparing the Docking Parameter File (DPF)

1. From the ADT GUI import receptor (PDBQT)

Docking>Macromolecule>Set Rigid Filename>cox2.pdbqt

2. From the ADT GUI import ligand (PDBQT)

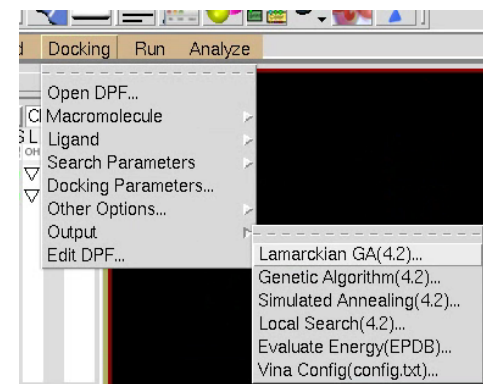
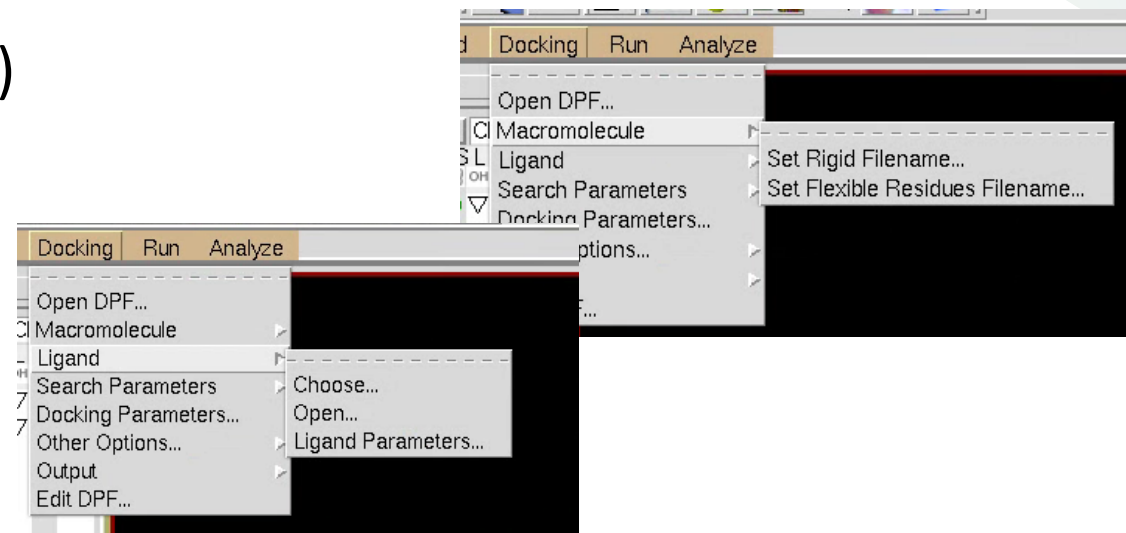
Docking>Ligand>Open>CEL.pdbqt

3. Save the Docking Parameter File (DPF)

Docking>Output>Lamarckian GA>docking-cox2.dpf

Alternatively you can run the following command

```
prepare_dp4.py -l CEL.pdbqt -r cox-2.pdbqt -o docking-cox2.dpf
```



Repeat the same procedure for COX1...



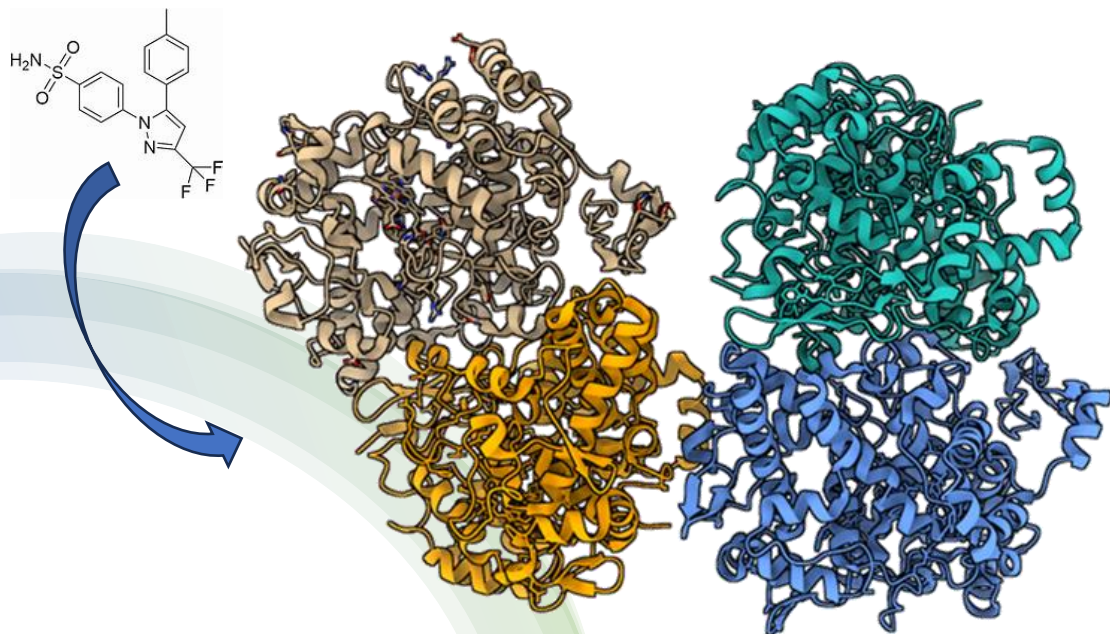
AutoDock 4

Molecular Docking Tutorial

RUN AutoDock4

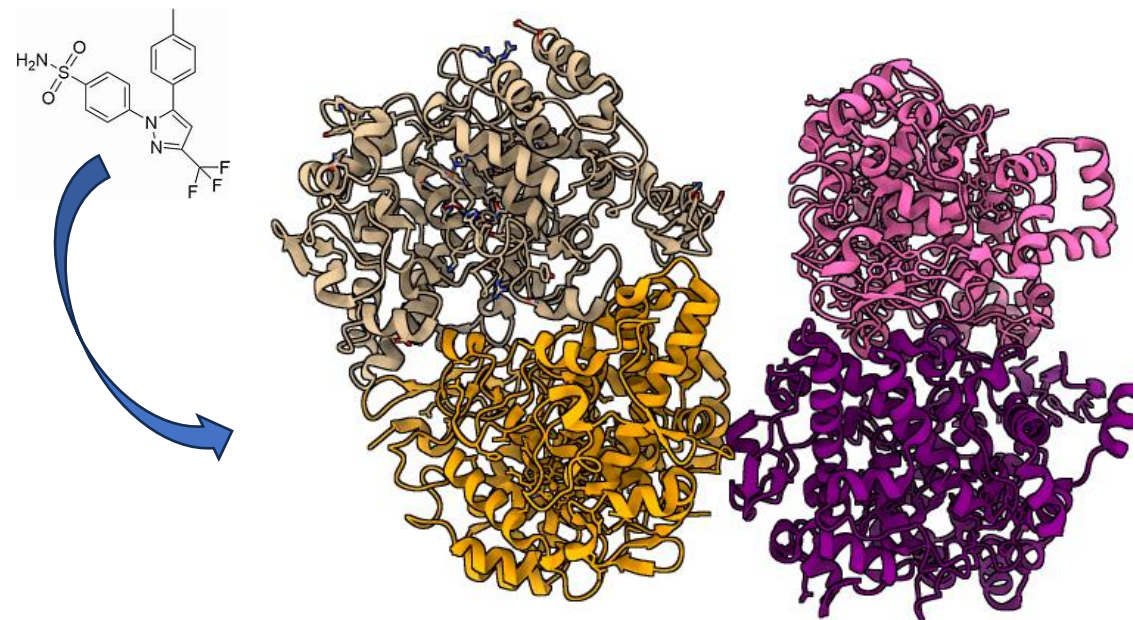
CEL vs COX2

```
Autodock4 -p docking-cox2.dpf -l docking-cox2.dlg
```



CEL vs COX1

```
Autodock4 -p docking-cox1.dpf -l docking-cox1.dlg
```



This will take around 4-5 minutes...

Molecular Docking Tutorial

Docking analysis (poses/scores)

1. Open the .dlg file

Analyze>Dockings>Open>file.dlg

2. Open the receptor (PDBQT)

Analyze>Macromolecule>Open>receptor.pdbqt

3. Visualize docking conformations

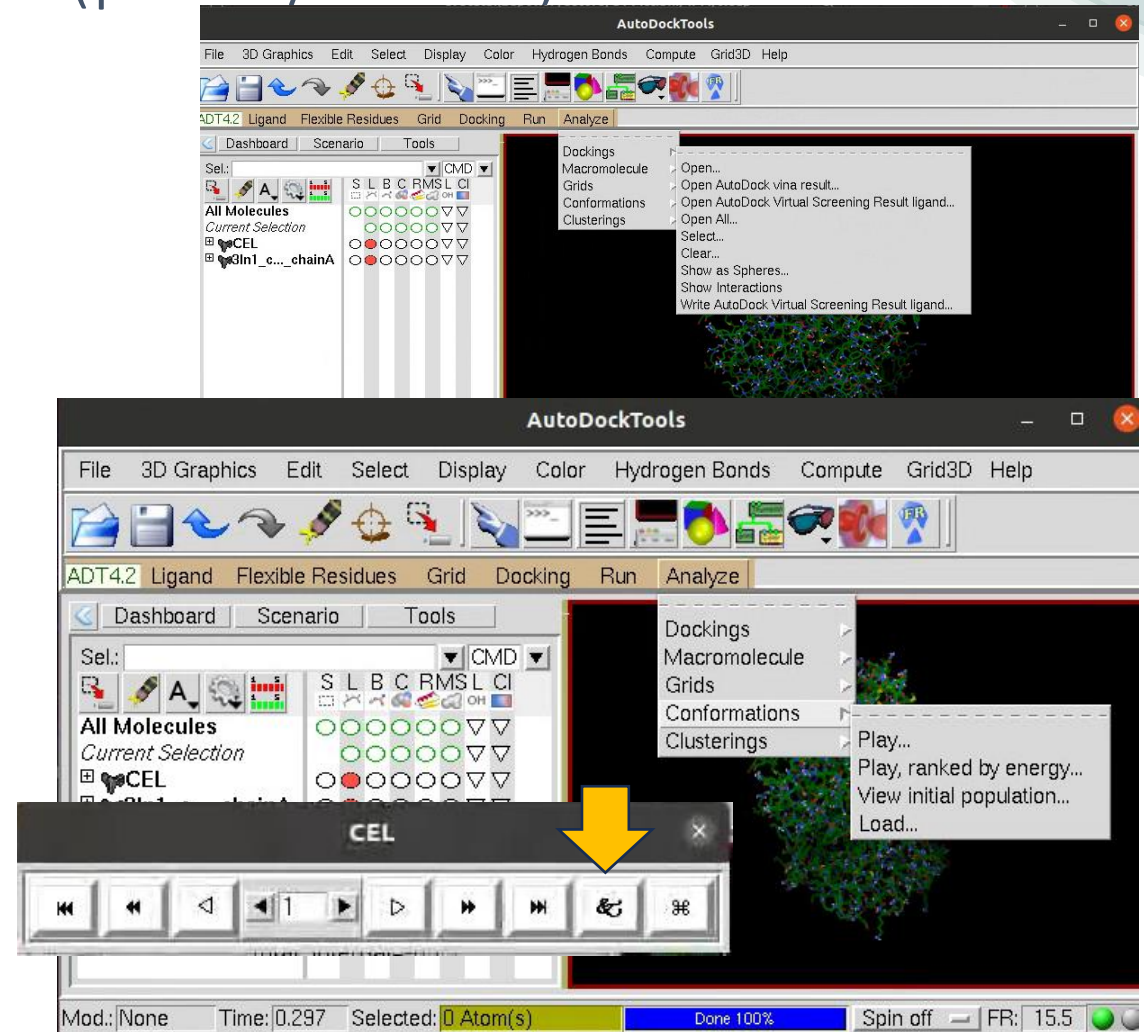
Analyze>Conformations>Play Ranked by Energy

4. Write all the conformations in pdbqt file

Click on the indicated icon>Write all

Docking conformations will be write ranked by energy.

«Conf0.pdbqt» has the best energy value



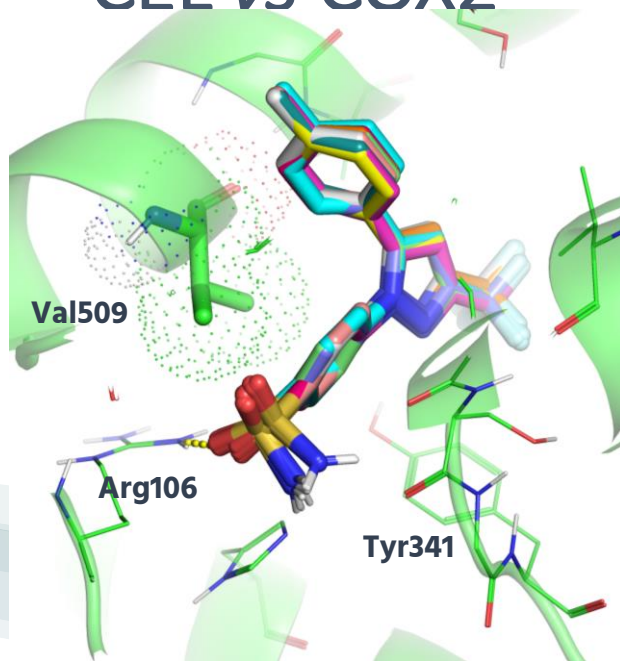


AutoDock 4

Molecular Docking Tutorial

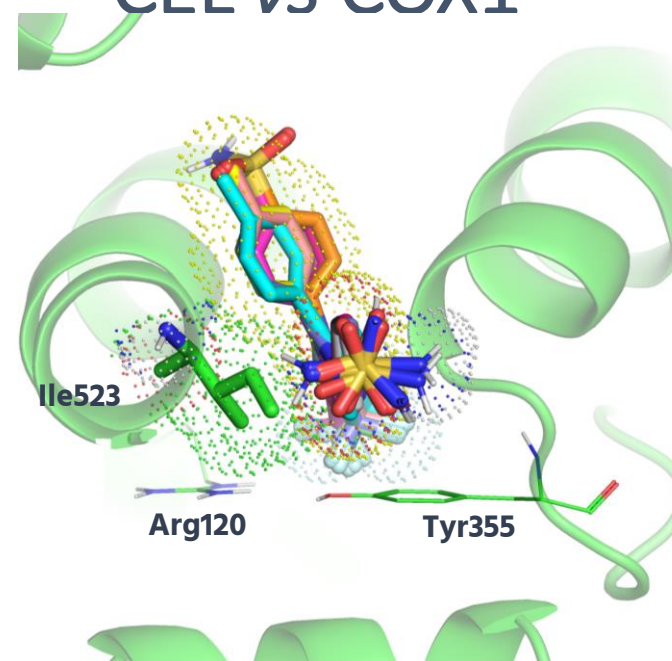
Docking analysis with PyMOL

CEL vs COX2



All the poses converge to one unique binding mode with a binding affinity score of -10.41 kcal/mol. The sulfonamide group directly interact with hydrogen-bonds with **Arg106**

CEL vs COX1



Of the 10 poses, 7 are oriented with the sulfonamide moiety toward **Arg120** and **Tyr355**, while 3 poses are in opposite orientation. Nonetheless, the presence of **Ile523** hampers Celecoxib to directly interact with **Arg120** and **Tyr355**. The binding affinity is also less stable (-7.62 Kcal/mol)



AutoDock Vina

Molecular Docking Tutorial



AutoDock Vina



Molecular Docking Tutorial

Docking parameter file (config.txt)

Protein
Ligand

```
receptor = cox2.pdbqt  
ligand   = CEL.pdbqt
```

Output file name

```
out      = vina_results.pdbqt  
log      = vina_results.log
```

Box center

```
center_x = 29.588  
center_y = -22.083  
center_z = -17.418
```

Box size

```
size_x   = 40  
size_y   = 40  
size_z   = 40
```

Search exhaustiveness

```
exhaustiveness = 8
```

Number of docking poses

```
num_modes      = 10
```

For an optimal comparison we will use the same grid coordinates and sizes as AutoDock4

Remember:

AutoDock Vina internally precalculates the grid maps. **You don't have to run AutoGrid4.**

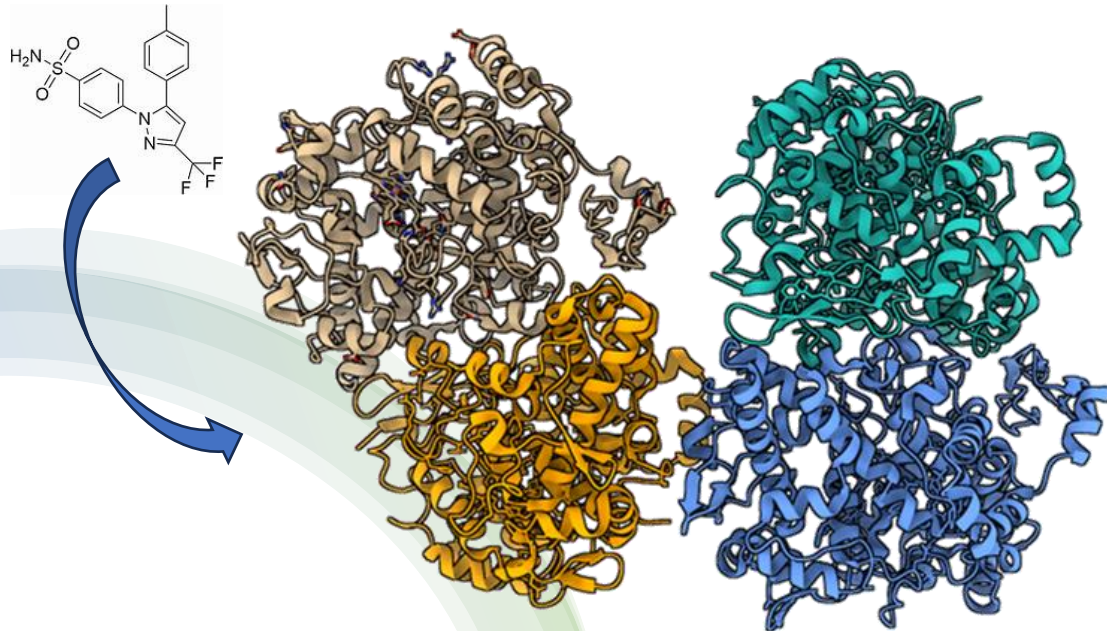


Molecular Docking Tutorial

RUN AutoDock Vina

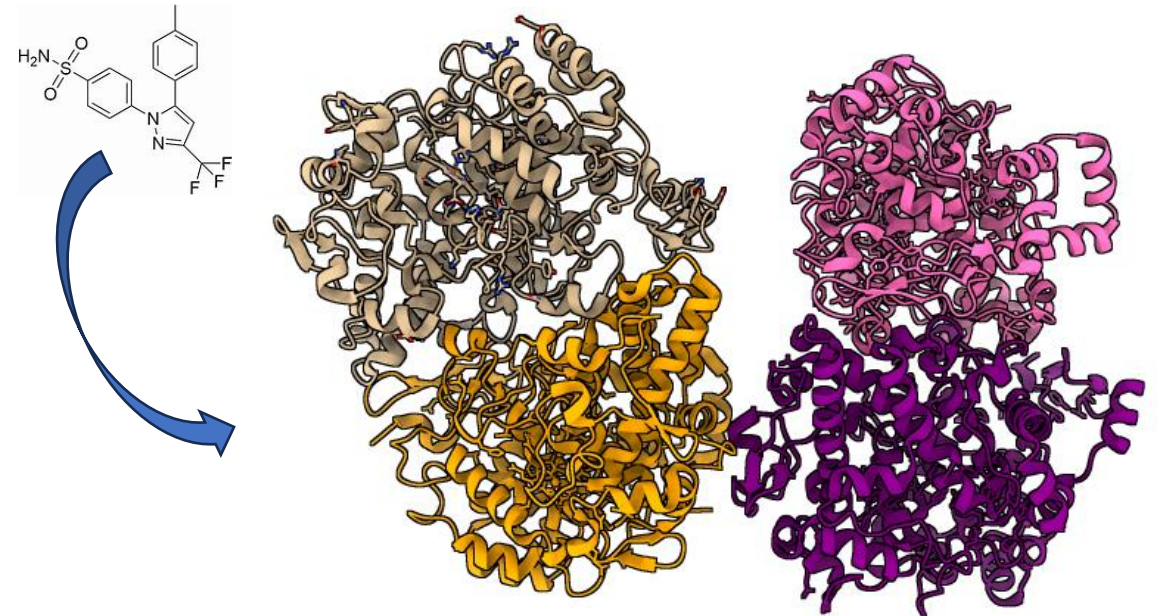
CEL vs COX2

```
vina --config config.txt
```



CEL vs COX1

```
vina --config config.txt
```



This will take around 1-2 minutes...

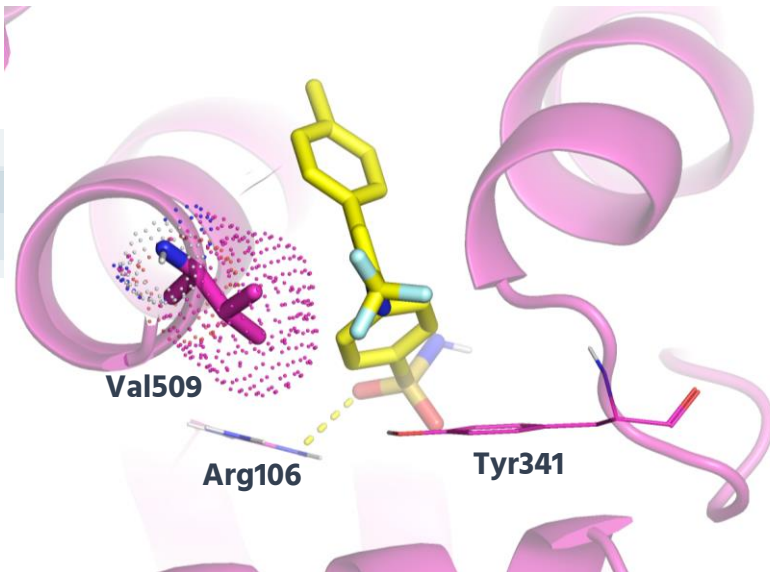


Molecular Docking Tutorial

Docking analysis with PyMOL

CEL vs COX2

1. Open the vina_results_cox2.pdbqt file and split
Action>State>Split
2. Open the cox2.pdbqt file



CEL vs COX1

1. Open the vina_results_cox1.pdbqt file
Action>State>Split
2. Open the cox1.pdbqt file

